MoS, Pixel Sensors for Optical Detection of Redox Molecules

CNF Project Number: 900-00

Principal Investigators: Paul L. McEuen^{1,2}, Jiwoong Park³, Daniel C. Ralph⁴ Users: Michael F. Reynolds¹, Marcos H.D. Guimarães^{1,2}, Hui Gao^{3,5}, Kibum Kang^{3,5}, Alejandro J. Cortese¹

Affiliations: 1. Laboratory of Atomic and Solid State Physics, 2. Kauli Institute at Cornell for Nanoscale Science,

Department of Chemistry and Chemical Biology, 4. Department of Physics; Cornell University, Ithaca NY:
Department of Chemistry, Institute for Molecular Engineering, and James Franck Institute,

University of Chicago, Chicago IL

Primary Sources of Research Funding: Cornell Center for Materials Research with funding from the NSF MRSEC program (DMR-1719875), Air Force Office of Scientific Research (AFSOR) multidisciplinary research program of the university research initiative Grant FA2386-13-1-4118

Contact: plm23@cornell.edu, mfr74@cornell.edu

Primary CNF Tools Used: Autostep i-line, ABM contact aligner, SC4500 evaporator, Oxford 81 etcher, VersaLine etcher

Abstract:

Spatially-resolved detection of redox molecules in solution is important for understanding chemical and biological systems. Optical detection is advantageously wire-free and easily multiplexed. We demonstrate that monolayer molybdenum disulfide (MoS₂) is a fast, sensitive, optical sensor for redox molecules.

Summary of Research:

Redox molecule detection has applications from detection of neurotransmitters in the brain to trace chemical detection in water samples. Traditional techniques, such as cyclic voltammetry, provide sensitive detection at a single electrode, but do not spatially resolve the variation in redox concentration. More advanced approaches including multiplexed electrode arrays [1,2] and numerous optical detection techniques [3-6] allow researchers to image redox molecules.

We demonstrate a wireless, optical approach for fast, sensitive redox imaging using a flexible, transferrable monolayer of MoS_2 . MoS_2 photoluminesces at about 650 nm [7], with an intensity that increases as the concentration of electrons on the MoS_2 decreases, as shown by back-gating [8] and chemical doping [9].

We use the doping dependence of MoS_2 photoluminescence (PL) to detect ferrocene/ferrocenium as a test redox couple. Metal-organic chemical vapor deposition MoS_2 samples [10], grown by Prof. Park's group, are patterned with a two-step fabrication process. First, we pattern contact pads on the MoS_2 with electronbeam evaporation. Second, we etch away the MoS_2 to define our device and pixel geometries, which are shown in Figure 1.

We performed two experiments to demonstrate that our MoS_2 pixel sensors measure the chemical potential of the solution. First, with a fixed total concentration of ferrocene/ferrocenium, we varied the ratio of the concentration of ferrocenium (Fc+) to ferrocene (Fc) in our solution while monitoring the PL of the MoS_2 (Figure 2A). The MoS_2 shows a marked increase in PL as Fc+/Fc increases. Second, in a solution without any ferrocene or ferrocenium, we apply a potential to the solution (denoted V_{LG} for liquid gate voltage) while grounding a contacted MoS_2 device. The PL is high at negative values for $V_{LG'}$ but decreases as V_{LG} is swept to positive values (Figure 2B, red curve). We compare the PL *versus* liquid gate voltage to PL *versus* change in chemical potential, where the change in chemical potential of the solution is given by

$$\Delta \mu = \frac{k_B T}{e} \ln \frac{[Fc]}{[Fc+]}$$

according to the Nernst equation (Figure 2B, blue dots). The good agreement between the two curves indicates that the PL of electrically floating pixels is set by the chemical potential of the solution.

Having characterized the MoS_2 sensors, we measure diffusion to demonstrate their speed and spatial resolution. We apply a voltage pulse to a microelectrode positioned above our MoS_2 pixel array in a solution of 1 mM ferrocene. The pulse oxidizes ferrocene to ferrocenium, which diffuses away from the probe tip, creating a spreading ferrocenium concentration that is imaged by the MoS_2 pixels (Figure 3). From these data, we extract a ferrocenium diffusion constant of about 1.8 × 10⁻⁹ m²/s, matching previous measurements [11].



Figure 1, left: A, Bright-field and B, PL (549 nm excitation, 650 emission) images of MoS₂ pixels and devices. *Figure 2, middle:* A, PL images of MoS₂ with varied ferrocenium/ferrocene ratios, showing increased PL with increased ferrocene concentration. B, PL versus liquid gate voltage for grounded device (red line) and versus solution chemical potential (blue dots), showing that MoS₂ sensors are Nernstian. Figure 4, right: A, Schematic and bright-field image showing PDMS microfluidic channel placed over MoS₂ pixel array. A pulse on the surface electrode oxidizes ferrocene to ferrocenium while a syringe pump flows the solution through the channel. B, MoS₂ PL imaging flow of ferrocene in the channel.



Figure 3: Frames from PL video imaging diffusion of ferrocenium away from a microelectrode after its potential is pulsed to 0.8 V versus the solution potential.

Finally, we use these sensors to image the flow of redox molecules in microfluidic channels. We mold polydimethylsiloxane (PDMS) microfluidic channels using silicon wafer masters patterned with the Plasma-Therm deep silicon etcher and place them over MoS₂ pixel arrays with platinum surface electrodes (Figure 4A). While flowing a ferrocene solution through the channel, we applied voltage pulses to the surface electrodes to oxidize ferrocene to ferrocenium and image laminar flow of the solution in MoS₂ PL (Figure 4B). These experiments show that MoS₂ can be used for real-time, spatially resolved imaging of redox molecules. The sensor could be improved by increasing the quantum efficiency of the MoS₂ PL in solution. In the future, these sensors could find applications in biological sensing experiments, e.g. spatially resolved detection of dopamine efflux from neurons.

References:

- J. Wang, R. Trouillon, Y. Lin, M. I. Svensson, A. G. Ewing, Individually Addressable Thin-Film Ultramicroelectrode Array for Spatial Measurements of Single Vesicle Release. Anal. Chem. 85, 5600–5608 (2013).
- [2] D. L. Bellin, et al., Electrochemical camera chip for simultaneous imaging of multiple metabolites in biofilms. Nat. Commun. 7, 10535 (2016).
- [3] X. Shan, U. Patel, S. Wang, R. Iglesias, N. Tao, Imaging local electrochemical current via surface plasmon resonance. Science. 327, 1363-6 (2010).
- [4] A. Jane, R. Dronov, A. Hodges, N. H. Voelcker, Porous silicon biosensors on the advance. Trends Biotechnol. 27, 230-239 (2009).
- [5] S. Kruss, et al., High-resolution imaging of cellular dopamine efflux using a fluorescent nanosensor array. Proc. Natl. Acad. Sci. U. S. A. 114, 1789-1794 (2017).
- [6] D. G. Hafeman, J. W. Parce, H. M. McConnell, Light-Addressable Potentiometric Sensor for Biochemical Systems (available at http://science.sciencemag.org/content/sci/240/4856/1182.full. pdf).
- [7] K. F. Mak, C. Lee, J. Hone, J. Shan, T. F. Heinz, Atomically Thin MoS₂: A New Direct-Gap Semiconductor. Phys. Rev. Lett. 105, 136805 (2010).
- [8] K. F. Mak, et al., Tightly bound trions in monolayer MoS₂. Nat. Mater. 12, 207-211 (2013).
- [9] S. Mouri, Y. Miyauchi, K. Matsuda, Tunable Photoluminescence of Monolayer MoS₂ via Chemical Doping. Nano Lett. 13, 5944-5948 (2013).
- [10] K. Kang, et al., High-mobility three-atom-thick semiconducting films with wafer-scale homogeneity. Nature. 520, 656-660 (2015).
- [11] Y. Wang, E. I. Rogers, R. G. Compton, The measurement of the diffusion coefficients of ferrocene and ferrocenium and their temperature dependence in acetonitrile using double potential step microdisk electrode chronoamperometry. J. Electroanal. Chem. 648, 15-19 (2010).